

Greener Journal of Pharmacy and Pharmacology

Anticancer Activity of *Carica papaya* Extracts in *vito* and Phytochemical Analysis

By

Khaled N. Rashed Gerda Fouche

Anticancer Activity of *Carica papaya* Extracts in *vito* and Phytochemical Analysis

Khaled N. Rashed^{1*}, Gerda Fouche²

¹National Research Centre, Pharmacognosy Department, Dokki, Giza, Egypt. ²Natural Product Chemistry Group, Council for Scientific and Industrial Research, PO Box 395, Pretoria 0001, South Africa.

*Corresponding Author's Email: khalednabih2015@yahoo.co.uk, Tel: 01003642233

ABSTRACT

The aim of the present work is the evaluation of anti cancer effect of *Carica papaya* aerial parts extracts and also the investigation of the phytoconstituents from the extracts of the plant. Petroleum ether (40-60°C), Chloroform, ethyl acetate and methanol 80% extracts of *C. papaya* aerial parts were tested for their anti cancer activity on three cancer cells TK10 (renal), UACC62 (melanoma) and MCF7 (breast) cancer cells using a Sulforhodamine B (SRB) assay. Petroleum ether of *C. papaya* at the concentration of 100 μ g/ml has shown a significant anticancer effect for MCF7 (breast) cancer cells and showed less anticancer effect for the other two cancer cells while the other extracts have mild anticancer effect on the three cancer cells. Phytochemical profile of the plant extracts proves the presence of flavonoids, tannins, alkaloids, carbohydrates and triterpenes and bioactive fractionation of petroleum ether extract revealed the isolation and identification of β -sitosterol, stigmasterol as the major constituents. The results suggest that petroleum ether extract of *C. papaya* exhibited a significant anti cancer activity for breast cancer.

Keywords: Carica papaya, aerial parts, anticancer activity, phytoconstituents.

INTRODUCTION

Cancer is a leading cause of death worldwide, it is a dreadful disease, and combating this disease is of great importance to public health. There is a necessity for search of new natural extracts of plant source and compounds with cytotoxic activity as the treatment of cancer with the available anticancer drugs is often unsatisfactory due to the problem cytotoxicity to the normal cells. Recent Phytochemical examination of plants which have a suitable history of use in folklore for the treatment of cancer has often resulted in the isolation of principles with anti cancer activity (Afolabi et al., 2007). Phytochemical examination has been making rapid progress and herbal products are becoming popular as sources of plausible anticancer compounds (Parag et al., 2010). In our searching for new anticancer drugs from plants, Carica papaya Linn, from Caricaceae family known as pawpaw tree is a vegetable fruit widely distributed throughout the world, mostly grow in tropics and it grows up to 5 to 10 m tall (Duke, 1984). The ripe fruit is edible and is usually eaten raw, without the skin or seeds. The unripe green fruit (which is a rich source of vitamin A) can be eaten cooked, usually in curries, salads and stews as used in Thai cuisine (Lohiya et al., 2002). The papaya fruit, as well as all other parts of the plant, contain a milky juice in which an active principle known as papain is present (Nisar et al., 2011). It has a good effect as a remedy in dyspepsia and kindred ailments. The juice has been in use on meat to make it tender (Nisar et al., 2011). The seed is used for intestinal worms when chewed. The root is chewed and the juice swallowed for cough, bronchitis, and other respiratory diseases. The unripe fruit is used as a remedy for ulcer and impotence (Aiyeloja and Bello, 2006). Previous studies on biological activities of C. papaya parts, extracts and isolated compounds showed that the latex and root extracts inhibited Candida albicans while extracts of pulp and seeds showed bacteriostatic properties against Staphylococcus aureus, Escherichia coli, Salmonella typhi, Bacillus subtilis, and Entamoeba histolytica, in vitro (Emeruwa, 1982). Its root aqueous extract has equally been shown to have purgative effect (Akah et al., 1997). In another study, the hypoglycemic and hypolipidemic effects of the aqueous seed extract of C. papaya was reported and the LD₅₀ of the oral toxicity was estimated to greater than 2000 mg/kg/oral route (Adeneye and Olagunju, 2009). This study was carried out to evaluate anti cancer effect of C. papaya extracts on three cancer cells TK10 (renal), UACC62 (melanoma) and MCF7 (breast) cancer cells and also to investigate the phytoconstituents of the plant extracts.

MATERIALS AND METHODS

Experimental

Spectroscopic data: NMR–Varian. MS (Finnigan MAT SSQ 7000, 70 ev). Silica gel (60-200 mesh, Merck). Thin Layer Chromatography (TLC): pre-coated sheets of silica gel 60 F₂₅₄ (Merck). Sephadex LH-20 (Sigma).

Plant Material

The aerial parts of *C. papaya* were collected from Al-Zohiriya garden, Giza, Egypt in May 2011. The plant was identified by Dr. Mohammed El-Gebaly, Department of Botany, National Research Centre (NRC) and by Mrs. Tereeza Labib Consultant of Plant Taxonomy at the Ministry of Agriculture and director of Orman botanical garden, Giza, Egypt. A voucher specimen is deposited in the herbarium of Al-Zohiriya garden, Giza, Egypt.

Preparation of the extracts

Finely ground aerial parts from *C. papaya* 700 g were extracted with petroleum ether (40-60°C) chloroform, ethyl acetate and methanol 80% solvents by maceration. Each extract was concentrated to dryness to yield, 9 g of petroleum ether, 6.7 g of chloroform, 5.5 g of ethyl acetate and 37 g of methanol 80% extracts, respectively. Each extract was tested for the presence of the phytoconstituents according to the following standard tests, Molisch's test for carbohydrates, Shinoda test for flavonoids, forth test for saponins, Salkowski 's for terpenes and sterols, FeCl₃ and Mayer's reagents for detecting of tannins and alkaloids, respectively (Sofowra 1993, Trease and Evans 1989, Harborne 1973).

Phytochemical characterization of Petroleum ether extract

Petroleum ether extract 7 g was subjected to silica gel column chromatography using petroleum ether as eluent and gradually increasing amount of ethyl acetate (EtOAc) where two major compounds were isolated. Compound 1 (β -sitosterol) was isolated from CHCl₃: EtOAc (90:10) elution and compound 2 (stigmasterol) was isolated through elution with petroleum ether: EtOAc (95:15).

Sulforhodamine B (SRB) assay

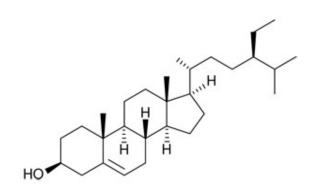
The growth inhibitory effects of the extracts were tested in the 3-cell line panel consisting of TK10 (renal). UACC62 (melanoma) and MCF7 (breast) cancer cells using a Sulforhodamine B (SRB) assay. The SRB assay was developed by (Skehan et al., 1990) to measure drug-induced cytotoxicity and cell proliferation. Its principle is based on the ability of the protein dye sulforhodamine B (Acid Red 52) to bind electrostatically in a pH-dependent manner to protein basic amino acid residues of trichloroacetic acid-fixed cells. Under mild acidic conditions, it binds to the fixed cellular protein, while under mild basic conditions it can be extracted from cells and solubilized for measurement. The SRB assay is performed at CSIR in accordance with the protocol of the Drug Evaluation Branch, NCI (National Cancer Institute). The 3-cell line panel used is recommended by the NCI for preliminary screens. The human cell lines TK10, UACC62 and MCF7 were previously obtained from NCI in the framework of a collaborative research program between CSIR and NCI. Cell lines were routinely maintained as monolayer cell cultures in RPMI containing 5% fetal bovine serum, 2 mM L-glutamine and 50µg/ml gentamicin. For the screening experiment, the cells (3-19 passages) were inoculated in 96-well microtiter plates at plating densities of 7-10 000 cells/well and were incubated for 24 h. After 24 h, one plate was fixed with TCA to represent a measurement of the cell population for each cell line at the time of drug addition (T0). The other plates with cells were treated with the experimental samples which were previously dissolved in DMSO as 10000µg/ml stocks and diluted in medium to a final concentration 100µg/ml. Cells without samples served as controls. Blank wells contained complete medium without cells. Emetine was used as a reference standard. The plates were incubated for 48 h after addition of the extracts (100 µg/ml). At the end of the incubation period, the cells were fixed to the bottom of each well with cold 50% trichloroacetic acid, washed, dried and dyed with SRB. Unbound dye was removed and protein-bound dye was extracted with 10mM Tris base for optical density determination at a wavelength 540 nm using a multiwell spectrophotometer. Optical density measurements were used to calculate the net percentage cell growth. The optical density of the test wells after 48-h period of exposure to test compound is Ti, the optical density at time zero is T0, and the control (untreated cells) optical density is C. Percentage cell growth is calculated as: [(Ti-T0)/(C-T0)] x 100 for concentrations at which Ti≥T0 [(Ti-T0)/T0] x 100 for concentrations at which Ti<T0.

RESULTS AND DISCUSSION

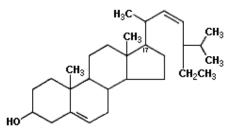
Phytochemical analysis of *C. papaya* extracts is shown in table 1, it showed that petroleum ether extract has triterpenes and/or sterols, chloroform extract has alkaloids and triterpenes, ethyl acetate extract has flavonoids and tannins while methanol 80% extract has flavonoids, tannins, alkaloids, carbohydrates and triterpenes. The isolated compounds from petroleum ether extract of *C. papaya* aerial parts are shown in Fig. 1.

Table 1: Phytochemical Analysis from the *C. papaya* aerial parts extracts

Constituents	Petroleum ether	Chloroform	Ethyl acetate	Methanol 80%	
Triterpenes and /or Sterols	+	+	-	+	
Carbohydrates and/or glycosides	-	-	-	+	
Flavonoids	-	-	+	+	
Coumarins	-	-	-	-	
Alkaloids and/or nitrogenous compounds	-	+	-	+	
Tannins	-	-	+	+	
Saponins	-	-	-	-	



Compound 1: β-sitosterol



Compound 2: Stigmasterol

Fig. 1: Compounds isolated from petroleum ether extract of C. papaya aerial parts

Structure elucidation of the bioactive compounds isolated of Petroleum ether extract of C. papaya

β-sitosterol (1): 20 mg, white needles, ¹H-NMR (400 MHz, CDCl₃): δ 5.37 (IH, m, H-6), 3.52 (IH, m, H-3), 1.09 (3H, s, CH₃-19), 0.98 (3H, d, *J*= 6.5, CH₃-21), 0.92 (3H, t, J= 7.4, CH₃-29), 0.85 (3H, d, J= 6.7Hz, CH₃-26), 0.81 (3H, d, J= 6.7Hz, CH₃-27), 0.75 (3H, s, CH₃-18). ¹³C-NMR(100 MHz, CDCl₃): δ 140.4 (C-5), 121.5 (C-6), 71.6 (C-3), 57.2 (C-17), 56.4 (C-14), 50.3 (C-9), 46.3 (C-24), 42,8 (C-13, 4), 39.8 (C-12), 37.6 (C-1), 36.7 (C-10), 35.9 (C-20), 34.2 (C-22), 31.7 (C-8, 7), 31.4 (C-2), 29.2 (C-25), 28.4 (C-16), 26.2 (C-23), 24.5 (C-15), 23.4 (C-28), 21.1 (C-11), 19.8 (C-26), 19.5 (C-19), 19.2 (C-27), 18.6 (C-21).

Stigmasterol (2): 17 mg, white needle crystals, ¹H-NMR (400 MHz, CDCl₃): δ 5.32 (IH, m, H-6), 5.11 (1H,dd, J= 14.2, 8.2 Hz, H-22), 5.04 (1H,dd, J= 14.2, 8.2 Hz, H-23), 3.54 (IH, m, H-3), 1.04 (3H, s, CH₃-10), 0.9 (3H, d, J= 6.5, CH3-20), 0.84 (3H, d, J= 7.4, CH₃-27), 0.82 (3H, d, J= 7.4, CH₃-26), 0.68 (3H, s, CH₃-13). ¹³C-NMR(100 MHz, CDCl₃): δ 140.6 (C-5), 138.4 (C-22), 129.1 (C-23), 121.8 (C-6), 71.9 (C-3), 56.7 (C-17), 56.9 (C-14), 50.9 (C-9), 50.7 (C-24), 42.6 (C-13, 4), 39.6 (C-12), 37.4 (C-1), 40.2 (C-20), 36.7 (C-10), 31.4 (C-8, 7), 31.7 (C-2), 30.9 (C-25), 28.8 (C-16), 24.8 (C-15), 24.7 (C-28), 21.5 (C-11), 20.8 (C-26), 20.4 (C-19), 19.7 (C-27), 19.1 (C-21).

Chromatographic separation and purification of the petroleum ether resulted in isolation and identification of compound 1 (β-sitosterol) which gave dark spot under short UV light that changed to violet colour on spraying with vanillin sulphuric and heating in an oven at 110 °C for 5min and also compound 2 (stigmasterol) gave a dark spot under short UV light and also gave a violet colour after spraving with vanillin sulphuric and heating in an oven at 110 °C for 5min. NMR spectral data has shown signals very close to compound 1 (β-sitosterol), also it is identified by other authors (Pateh et al. 2009). Similarly, a comparison of the NMR spectral data of compound 2 (Pateh et al. 2009; Shirin et al., 2012) with published data allowed the identification of compound 2 as stigmasterol; confirmation of both compounds was done by co-TLC with authentic standards. The extracts of C. papaya aerial parts were tested for their anticancer activities on different cancer cells, TK10 (renal), UACC62 (melanoma) and MCF7 (breast) cancer cells using Sulforhodamine B (SRB) assay (Skehan et al., 1990). Petroleum ether (40-60°C), Chloroform, ethyl acetate and methanol 80% extracts of C. papaya aerial parts have a mild anticancer effect for TK10 (renal) and UACC62 (melanoma) cancer cells while petroleum ether has a significant anticancer effect for MCF7 (breast) cancer cells with comparison with a standard drug (Emetine) and the other extracts showed a mild anticancer effect for this cancer cell (table 2). The significant anticancer effect of petroleum ether extract of C. papaya is may be due the presence of the bioactive isolated compounds, βsitosterol and stigmasterol from the extract. In previous study, Chorisia crispiflora n-hexane extract has a significant effect on growth inhibition and induction of apoptotic processes in MCF-7 breast cancer cells and the activity is due to the presence of β-sitosterol, β-sitosterol 3-glucoside and stigmasterol 3-glucoside (Samar et al., 2013) and in another reported study that β -sitosterol increased the activities of antioxidant enzymes, superoxide dismutase and glutathione peroxidase in cultured macrophage cells with oxidative stress induced by phorbol 12myristate 13-acetate, indicating that phytosterols can protect cells from damage by reactive oxygen species (Vivancos and Moreno, 2005).

Extracts	Con., μg/ml	Growth TK10, %	SD	Growth UACC62, %	SD	Growth MCF7,%	SD
Petroleum ether							
extract	100	-25.54	0.037	-42.17	0.059	-62.91	0.080
Chloroform	100	88.10	0.019	48.08	0.042	53.89	0.091
Ethyl acetate	100	74.23	0.012	40.89	0.016	33.68	0.020
Methanol 80%	100	-18.69	0.001	-19.32	0.071	-29.06	0.021
EMETINE	10µM	-61.35	0.007	-86.66	0.006	-46.41	-61.35

SD is standard deviation. % Growth is the net growth of the cells in treated wells compared to untreated controls over the 48h experimental period, i.e. 100% growth means there are the same amount of cells in treated wells as in untreated control wells; 0% growth means the treated wells contain the same number of cells as at the start of the incubation, time 0 (thus no increase in cell number); -100% growth means there are no cells left in the well after 48h. Emetine as a standard.

CONCLUSION

The reported results show that petroleum extract of *C. papaya* aerial parts has significant anticancer effect on MCF7 (breast) cancer cells. Thus, *C. papaya* aerial parts could be helpful in cancer prevention and treatment. *C. papaya* could be a natural source of anticancer compounds with anti proliferative and/or apoptotic properties and as well, due to its anticancer pharmacological effect, clinical trials are recommended to evaluate the beneficial effects of this plant in human models.

Conflict of interest

There is no conflict of interest associated with the authors of this paper.

REFERENCES

- Adeneye A.A. and Olagunju J.A. 2009. Preliminary hypoglycemic and hypolipidemic activities of the aqueous seed extract of *Carica papaya* Linn. in Wistar rats. Biology and Medicine 1(1): 1-10.
- Afolabi C.A., Ibukun E.O., Emmanuel A., Obutor E.M. and Farombi E. O. 2007. Phytochemical Constituent and antioxidant activity of extract from leaves of *Ocimium Gratissium*. Science Research Essay 2(5):163-166.
- Aiyeloja A. and Bello O.A. 2006. Ethnobotanical potentials of common herbs in Nigeria: A case study of Enugu state. Educational Research and Review 1 (1):16-22.
- Akah P.A., Oli A.N., Enwerem N.M. and Gamaniel K. 1997. Preliminary studies on purgative effect of *Carica papaya* root extract. Fitoterapia 68(4): 327-331.
- Duke J.A. 1984. Borderline herbs. Boca Raton: CRC Press.
- Emeruwa A.C. 1982. Antibacterial substance from *Carica papaya* fruit extract. Journal of Natural Products 45(2):123-127.
- Harborne J.B. 1973. Phytochemical Methods. Chapman and Hall. Ltd., London, 49-188.
- Lohiya N.K., Manivanna B., Mishra P.K., Pathak N., Sriram S., Bhande S.S., Panneerdoss S. 2002. Chloroform extract of *Carica papaya* seeds induce long-term reversible azoospermia in Langur monkey. Asian Journal of Andrology 4: 17-26.
- Parag R. P., Bhuvan P. R. and Hamsraj A. K. 2010. Potent antitumor activity of *Rubia cordifolia*, International Journal of Phytomedicine 2:44-46
- Pateh U.U., Haruna A.K., Garba M., Iliya I.M., Abubakar M.S. and Ambi A.A. 2009. Isolation of Stigmasterol, β-Sitosterol and 2-Hydroxyhexadecanoic acid methyl ester from the Rhizomes of Stylochiton lancifolius and Kotchy (Araceae). Nigerian Journal of Pharmaceutical Sciences 7 (1):19-25.
- Nisar A., Hina F., Muhammad A., Bilal H. A., Ijaz M. and Lubna F. D. 2011. fever treatment with *Carica papaya* leaves extracts. Asian Pacific J Tropical Biomedicine 12:330-333.
- Sofowra A. 1993. Medicinal Plants And traditional Medicine in Africa. Spectrum Books Ltd., Ibadan, Nigeria, 191-289.
- Trease G.E., Evans W.C. 1989. Pharmacology, 11th edn., Bailliere Tindall, London, 45-50.
- Samar S. A., Abeer M. A. and Omayma A. E. 2013. Phytochemical Investigation and Molecular Profiling by p21 and NF-κB of *Chorisia crispiflora* Hexane Extract in Human Breast Cancer Cells *in* Vitro. British Journal of Pharmaceutical Research 3(1):78-89.
- Shirin M., Abdul Majid A., Mustafa G., Mohammed R.M., Masoud R., Mohsen S. 2012. Phytochemical analysi and Metal-chelation activity of *Achillea tenufolia* Lam. Iranian Journal of Pharmaceutical Research 11 (1):177-183.
- Skehan, P., Storeng R., Scudiero D., Monks A. and McMahon J. 1990. New colorimetric cytotoxicity assay for anticancerdrug screening. Journal National Cancer Institute 82: 1107-1112.
- Vivancos M. and Moreno J.J. 2005. beta-Sitosterol modulates antioxidant enzyme response in RAW 264 7 macrophages. Free Radic Biol Med 39:91–97.